## Electron Microscope preparation

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	Transmission (TEM) & Scanning(SEM) &Scanning Transmission (STEM)	Cryo-Scanning (Cryo-SEM) & (Cryo- STEM)	Graphene Liquid Cell Electron Microscopy (GLC-EM)
USED FOR:	Biological	Biological	Biological (that cannot withstand a vacuum)
PREP PRIOR TO	"Purification" and possible staining	"Purification", <b>Negative stain</b> (*see below) and/or Crosslinking solution (glycerol or sucrose solution)	
FIXING	Chemical - to cross link proteins to other molecules	Cryo - Rapid freezing (prep is -4C, -90C for sectioning)	None
Chemicals used	Aldehydes (common one used is gluteraldehyde); Familiar one is Formaldehyde. Acrolein Tannic Acid	Liquid Helium Liquid Nitrogen	
RINSING (needed for reducing acidity bc of fixing)	sodium cacodylate	X	
SECONDARY	To further stabilize the the tinier structures in the	X	
FIXING	specimens and Lipids		
Chemicals used	Osmium tetroxide OsO4 Potassiumferrocyanide. Picric acid (for osmium-free fixing)		
DEHYDRATION	Removes the water and replaces it with solvent	X	NONE
Chemicals used	Ethanol or Acetone followed by Hexamethyldisilazane (HMDS) or liquid CO2		
INFILTRATION	For slicing and polymerization (60C), overnight	X	Liquid mediums containing gold or
			other metal nanoparticles for "labeling".
Chemicals used	Propylene oxide Epon or Epon-Araldite Acetone Acrylic Resin		Metals like GOLD; Fluorescent quantum dot nanoparticles
STAINING (negative)	Biological samples usually need staining for contrast (creating dark particles on a light background)	Done prior to Fixing (freezing)	
STAINING (positive)	Biological samples are stained to see contrast (light particles on a dark background)		
Chemicals used	Uranium (uranyl acetate, uranyl formate) Lead (lead citrate, Lead hydroxide) Tungsten (phosphotungstic acid) Molybdenum (Ammonium Molybdate) Osmium (osmium ferricyanide, osmium tetroxide) Gold (auroglucothionate)		
SPURRING	X	Vapor shadowing to help with contrast	
Chemicals used		platinum gold carbon	